

Rethinking Antimicrobial Effectiveness Testing: Scientific Best Practices for USP 51 Compliance

USP <51> Antimicrobial Effectiveness Testing (AET) is a critical assay for validating the efficacy of preservatives in aqueous-based pharmaceutical and personal care products. While the test is widely recognized by global regulatory bodies, its reliability is contingent on factors often overlooked in basic implementation— neutralization validation, media suitability, and regulatory documentation. This white paper outlines scientific best practices for executing USP <51> in a way that ensures data integrity and supports product development across the R&D to commercialization continuum.

Introduction

The increasing complexity of formulations in the pharmaceutical and personal care industries has heightened the importance of rigorous microbial control testing. USP <51>, established by the United States Pharmacopeia, provides a robust framework to assess whether a product's preservative system effectively controls microbial growth throughout its shelf life. Despite its prevalence, many manufacturers treat AET as a binary hurdle, failing to extract critical formulation insights or anticipate compliance risks.

This paper evaluates the assay's core requirements, common execution challenges, and how a scientifically integrated approach—such as that used at iFyber—can yield more actionable results and de-risk product development.

Test Overview and Requirements

Product Categories

USP has divided product types into 4 different categories, refer to **Table 1**. The category determines the acceptance criteria and parameters of the study.

Table 1. USP categories for products.

Category	Product Description	
1	Injections, other parenterals including emulsions, otic products, sterile nasal products, and ophthalmic products made with aqueous bases or vehicles.	
2	Topically used products made with aqueous bases or vehicles, nonsterile nasal products, and emulsions, including those applied to mucous membranes.	
3	Oral products other than antacids, made with aqueous bases or vehicles.	
4	Antacids made with an aqueous base	

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Required Species

The product is tested against 5 different species of microorganisms:

- Escherichia coli ATCC 8739
- Pseudomonas aeruginosa ATCC 9027
- Staphylococcus aureus ATCC 6538
- Candida albicans ATCC 10231
- Aspergillus brasiliensis ATCC 16404

Testing of Products

On the first day of testing or initial testing, the sample is measured out into 5 equal aliquots. For categories 1,2, and 3, between $1x10^5$ to $1x10^6$ cfu/mL of each microorganism is added to one of the aliquots. For category 4, the samples are inoculated with $1x10^3$ to $1x10^4$ cfu/mL of inoculum. The samples are then plated on appropriate agar and kept at appropriate conditions for the given species. The plates are then counted as log(cfu/mL). The remaining inoculated samples are stored at 22.5 ± 2.5 °C. The samples are then plated again on day 7, 14 and/or 28, depending on sample type. The calculated results are then compared against each time point. Acceptance criteria are based on microorganism type and product type as shown in **Table 2**.

Acceptance Criteria

The counts are calculated as log/CFU, but results are typically reported as pass/fail. **Table 2** lists the criteria for bacteria and yeast/mold for each product category. For a product to pass, it must have satisfactory results for all five species.

Category	Criteria for Bacteria	Criteria for Yeast and Molds
١	 Not less than 1.0 log reduction from the initial calculated count at 7 days Not less than 3.0 log reduction from the initial count at 14 days No increase from the 14 days' count at 28 days. 	 No increase from the initial calculated count at: 7 14 and 28 days.
2	 Not less than 2.0 log reduction from the initial count at 14 days, No increase from the 14 days' count at 28 days. 	 No increase from the initial calculated count at: 14 and 28 days.
3	 Not less than 1.0 log reduction from the initial count at 14 days, No increase from the 14 days' count at 28 days. 	 No increase from the initial calculated count at: 14 and 28 days.
4	• No increase from the initial calculated count at 14 and 28 days.	 No increase from the initial calculated count at: 14 and 28 days.

Table 2. Acceptance criteria for each product category.







Additional Requirements/Testing for USP

- Sterility checks No microbial growth in a batch of media.
- Growth promotion The media properly grows microorganisms of interest.
- Suitability Testing Checks for proper neutralization.

Why are these additional requirements necessary?

- Sterility and Growth Promotion Important quality checks that confirms the usability of the media.
- Suitability Testing Every product has unique properties that can affect the outcome of results. Proper neutralization ensures that any antimicrobial properties are inactivated so accurate counting of microbes can occur. Improper neutralization can lead to false negatives and incorrect results. It is highly recommended that a neutralization validation study is performed prior to starting a USP 51 assay, this will help prevent inaccurate results.

Case Examples from iFyber

- **Topical Gel with Rebound at Day 28**: iFyber helped reformulate the preservative system and validated a new neutralizer. The client met FDA submission timelines with no additional delay.
- Oral Suspension with Failed Yeast Reduction: Suitability testing revealed an excipient was interfering with recovery. A protocol modification resolved the issue.

Conclusion

USP <51> remains a gold-standard method for antimicrobial effectiveness testing, but its value is maximized only when executed with scientific rigor. Laboratories that treat AET as a checkbox risk missing formulation insights, regulatory readiness, and even patient safety considerations. iFyber's approach blends microbiological expertise with regulatory foresight, offering clients a more strategic pathway through preservative validation and beyond.

To learn more about iFyber's approach to antimicrobial testing or to initiate a project, visit <u>www.ifyber.com/connect</u>.



